

## Protocol for in vitro propagation of HUVEC/TERT2 cells

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Reagents
<p>Gelatin solution (Sigma-Aldrich, Cat# G1393) PBS Trypsin-EDTA (Gibco, Cat# 25300054) EndoUp2 (Evercyte, Cat# MHT-006-2) which contains:</p> <ul style="list-style-type: none"> <li>• EBM™ Basal Medium 500 mL (Lonza Cat# CC-3121) with selected supplements from EGM™ SingleQuots™ Kit (Lonza, Cat# CC-4133) – namely BBE (bovine brain extract), HEGF, hydrocortisone solution and ascorbic acid solution</li> <li>• 10 % Fetal Bovine Serum (FBS) (Sigma-Aldrich, Cat# F7524)</li> <li>• 20 µg/ml G418 (InvivoGen, Cat# ant-gn-5)</li> </ul>
Practical application
<p>The culture flasks have to be pre-coated with gelatin (diluted to 0.1 % in PBS). Therefore, the culture flasks are treated with gelatin solution (60 µl/cm<sup>2</sup>) at 37°C for at least 10 min (10 – 60 min). Before introducing cells, remove excess of gelatin solution and rinse flask once with PBS (160 µl/cm<sup>2</sup>).</p> <p>For detachment of cells remove and discard culture medium and wash cells once with PBS. Remove PBS completely. Then, add 0.05 % Trypsin-EDTA (1x) solution (room-temperature; 20 µl/cm<sup>2</sup>), make sure that all cells have been in contact with this solution and incubate the culture flask at 37°C for approximately 2 – 3 min. Observe cell detachment under an inverted microscope. As soon as all cells are detached (if necessary agitate the cells by gently hitting the flask), add growth medium (about 160 µl/cm<sup>2</sup>) and aspirate cells by pipetting. Add appropriate aliquots of the cell suspension to gelatin coated culture vessels supplemented with growth medium (final volume of 240 µl/cm<sup>2</sup>).</p> <p>Cells are grown in EndoUp2 ready-to-use medium at 37°C in a humidified atmosphere with 5 % CO<sub>2</sub>. A split ratio of 1:4 to 1:6 twice a week is recommended (after having reached about 90 – 95 % confluence).</p>
Related products
<ul style="list-style-type: none"> <li>• EndoUp2 ready-to-use medium, 500 ml (Evercyte, Cat# MHT-006-2)</li> </ul>